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Toxicological Evaluation of Aqueous Extract of *Khaya senegalensis* Stem Bark on Liver function Indices in Albino Rats

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ABSTRACT

Toxicological evaluation of aqueous extract of Khaya senegalensis stem bark on some liver function parameters of normal rats (100-140g) were critically examined. The albino rats (16) were randomly assigned into four (I-IV) groups each of which contains four rats. They were acclimatised for a week and Khaya senegalensis stem bark extract was administered for three weeks after which they were sacrificed. Group I (Control) received equivalent volume of distilled water while group II, III and IV received 11.62 mg/kg bwt, 13.95 mg/kg bwt and 16.28 mg/kg bwt of the extract respectively. The activities of ALT, AST and ALP in serum at all doses of the extract was significantly ($p < 0.05$) increased compared with the control. Also, the concentration of total protein and globulin in the serum was significantly ($p < 0.05$) increased while the concentration of albumin in the serum was significantly ($p < 0.05$) decreased when compared with the control. Overall, the results indicated that oral administration of aqueous extract of Khaya senegalensis stem bark once daily for three weeks at the doses of 11.62, 13.95 and 16.28 mg/kg body weight caused hepatocellular toxicity and could hamper normal functioning of the liver of the animals. Therefore, the aqueous extract of Khaya senegalensis stem bark is not safe as an oral remedy at the doses investigated in the present study.

Key words: *Khaya senegalensis, Stem bark, Liver function parameters, Medicinal herbs.*

INTRODUCTION

The use of medicinal herbs in traditional system of medicine is a common practice in many cultures around the world, especially in African society. This practice has gained widespread acceptance in developing as well as in developed nations. Researchers are also beginning to appreciate the role of medicinal plants in health care delivery. This is as a result of the effectiveness, low cost and the availability of these herbal medicines¹. It is noteworthy that some orthodox medicines in use today were developed from the biochemical templates obtained from medicinal plants. However, the widespread use and popularity of herbal medicines do not guarantee their efficacy and safety². Therefore, there is need for detailed scientific analyses and adequate information on the toxicity of commonly used herbal drugs³. The way to determine the safe or unsafe use of a medicinal plant is the assessment of how it affects hematological and biochemical parameters^{4,5}. Changes from normal physiological levels of these parameters after administration of a chemical agent to the experimental animals is an indication of adverse effects of such agent on living organisms⁶.

Khaya senegalensis (Ders.) A. Juss is a large and sturdy tree (up to 35m high with a diameter of 1 to 3m) of Meliaceae family. Also named Senegal mahogany, it is a forestry species well known and exploited by Africans⁷. Phytochemical screening of trunk bark allowed Lompo⁸ to highlight the main chemical groups

in *K. Senegalensis*: fatty acids, carotenoids, coumarins, emodols, tannins, compounds reducers, anthracenosides, steroidal glycosides, flavonosides, carbohydrates, saponins, sterols and triterpenes, anthocyanins. Recently, Yuan⁹ and Bickii¹⁰ reported the isolation of some limonoids named Khayalenooids from the stem and bark. Yuan⁹ elucidated the structures of those molecules based on spectroscopic analysis. The stem bark extract is used for treating jaundice, malaria, dermatoses and hookworm infections¹¹. Limonoids isolated from other one species of *Khaya* (*Khaya grandifoliola*) is declared highly effective against the causative agent of malaria, *Plasmodium falciparum*¹⁰. Also, It has been reported that the plant is used in the treatment of Diarrhoea¹², Bacterial Infections¹³, Cancer^{14,15}, Helminthosis^{16,17}, Trypanosomosis¹⁸, Diabetes^{19,20,21,22}, mental illness²³. Many of the indigenous plants are used by man without the actual knowledge of their toxic potentials in an attempt to cure diseases and relief physical suffering²⁴. The present study was therefore undertaken to determine the effect of aqueous extract of *Khaya senegalensis* stem bark on liver function parameters in normal rats.

MATERIALS AND METHOD

Plant Material

The stem barks of *Khaya senegalensis* were obtained from Bayero University Kano, old Campus and was authenticated at the Herbarium of the Department of Plant Biology, Bayero University Kano, Nigeria, where a voucher specimen was deposited at the Herbarium of the Institute.

Experimental Animals

Wister male and female adult albino rats (16) weighing between 100-140g were obtained from National Veterinary Research Institute, Vom, Jos, Nigeria. The animals were housed in aluminum cages under standard conditions. They were maintained on standard animal pellets and water *ad libitum*. The animals were acclimatized for two weeks before the commencement of the experiment.

Chemicals and reagents

The assay kits for alkaline phosphatase, aspartate and alanine transaminases were products of Randox Laboratories, United Kingdom. All other reagents used were of analytical grade and were prepared in glass distilled water.

Preparation of Plant Extract

The stem barks of *Khaya senegalensis* were oven dried at 40°C for 72 hours to a constant weight. The dried stem barks were then pulverized using Beltone Luinohun Blender (model MS-223, Taipei, Taiwan). The powdered material was stocked in a plastic container from which 100 g was extracted in 1000 ml of cold distilled water for 48 hours at 37°C. This was then filtered with Whatman No. 1 filter paper. The filtrate was concentrated on a steam bath. The extract was reconstituted in distilled water to give the required doses of 11.62, 13.95 and 16.28 mg/kg body weight as used in this study. The reconstituted aqueous extract was administered orally using cannula to all the animals in different groups²⁵.

Experimental Design

The albino rats were divided into four groups of four rats each. The normal dose of *Khaya senegalensis* for average weight human being is 8140 mg/70kg body weight which is equivalent to 116.3 mg/kg body weight. Group I served as control, group II, III, IV were treated with 11.62, 13.95 and 16.28 mg/kg body weight of aqueous extract of *K. senegalensis* for three weeks.

Collection of blood sample and Preparation of serum

The rats were placed under diethyl ether anaesthesia; the neck area was quickly shaved to expose the jugular veins. The veins after being slightly displaced (to avoid contamination with interstitial fluid) were then cut with a sterile scalpel blade. Blood samples were then collected into clean dry centrifuge tubes and were allowed to clot for 30 minutes. This was then centrifuged at 33.5 g for 15 minutes using a Uniscope Laboratory Centrifuge (model SM800B). The sera were aspirated with Pasteur pipettes and stored frozen overnight at -20°C before being used for the biochemical analyses²⁵.

Determination of biochemical parameters

Total protein was determined using the Biuret method of Henry *et al.*²⁶ Activities of aspartate transaminase (AST) (E.C.2.6.1.1) and alanine transaminase (ALT) (2E.C.2.6.1.2) were determined based

on the method described by Schmidt and Schmidt²⁷ while alkaline phosphatase (ALP) (E.C.3.1.3.1) activity was determined as described by Wright *et al.*²⁸ The concentration of albumin was determined as described by Grant and Kacchman²⁹. All measurements were done using Spectronic 21 spectrophotometer (Bausch and Lomb, NY).

Statistical analysis

The data were expressed as mean \pm standard deviation (SD). Statistical analysis was performed using analysis of variance (ANOVA) and Duncan multiple range test at 5% level of confidence ($p < 0.05$).

RESULTS

Table 1 shows the activities of aspartate transaminase (AST), alanine transaminase (ALT) and alkaline phosphatase (ALP) in the serum of rats administered with aqueous extract of *khaya senegalensis* stem bark. There was a significant ($p < 0.05$) increase in the serum activities of ALT, AST and ALP at all doses of the aqueous extract of *khaya senegalensis* when compared with the control group. Also, the increase was shown to be dose dependent at all treated groups when compared with the control (Table 1).

Table 2 shows the concentration of total protein, albumin and globulins in the serum of rats administered with aqueous extract of *khaya senegalensis* stem bark. There was a significant ($p < 0.05$) increase in the serum concentration of total protein and globulins at all doses of the extract while there was a significant ($p < 0.05$) decrease in the concentration of serum albumin when compared with the control. Also, the increase in the concentration of serum total protein, globulins and decrease in the concentration of serum albumin was shown to be dose dependent (Table 2).

Table 1: Activity of ALT, AST and ALP in serum of rats administered with aqueous extract of *Khaya senegalensis* stem bark

Group	ALT (U/L)	AST(U/L)	ALP (U/L)
Control	2.00 \pm 0.00 ^a	13.00 \pm 2.69 ^a	27.75 \pm 2.25 ^a
11.52 mg/kg bwt	5.00 \pm 0.00 ^d	15.25 \pm 5.89 ^d	29.50 \pm 1.66 ^d
13.95 mg/kg bwt	6.50 \pm 0.87 ^c	18.50 \pm 3.57 ^c	33.54 \pm 5.89 ^c
16.28 mg/kg bwt	8.75 \pm 1.09 ^b	20.25 \pm 4.09 ^b	45.00 \pm 3.67 ^b

Values are expressed as Mean \pm SD (n = 4). Values in each column with different superscript (a-d) are significantly different ($P < 0.05$). ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase.

Table 2: Concentration of Total protein, Albumin and Globulins in rats administered with aqueous extract of *Khaya senegalensis* stem bark

Group	Total protein (g/L)	Albumin (g/L)	Globulins (g/L)
Control	63.25 \pm 2.04 ^a	38.25 \pm 1.48 ^a	25.75 \pm 1.48 ^a
11.52 mg/kg bwt	67.00 \pm 1.00 ^c	34.75 \pm 0.43 ^a	29.25 \pm 1.30 ^c
13.95 mg/kg bwt	70.75 \pm 2.68 ^c	33.50 \pm 1.69 ^a	33.25 \pm 3.24 ^d
16.28 mg/kg bwt	73.00 \pm 2.45 ^b	30.25 \pm 3.83 ^a	36.75 \pm 1.30 ^b

Values are expressed as Mean \pm SD (n = 4). Values in each column with different superscript (a-d) are significantly different ($P < 0.05$).

DISCUSSION

Most reports on toxic effects as a result of the use of herbal medicines and dietary supplements are associated with hepatotoxicity, although untoward effects on other organs such as kidney, skin, brain and the heart have been published^{30,31}.

The aminotransferases (ALT and AST) are 'markers' of liver damage and can thus be used to assess liver cytolysis with ALT being a more sensitive biomarker of hepatotoxicity than AST³². The increase in the activity of serum ALT and AST as observed in this study might be a sign of hepatocellular damage^{33,34,35}.

ALT and AST are located in the cytoplasm and mitochondria of liver cells in high concentrations but low in blood. However, ALT is more liver-specific³⁶. It is known that increased activities of these enzymes in serum are due to increased membrane permeability and leakage into the blood circulation when there is damage to liver cells³⁵. Thus, rise in the activities of ALT and AST in the serum due to hepatic necrosis may be noticed several days before clinical signs are manifested³⁷. Alkaline phosphatase (ALP) is a 'marker' enzyme of damage for the plasma membrane and endoplasmic reticulum^{38,39}. It is frequently used to assess the integrity of the plasma membrane⁴⁰. In this study, there was an increase in the activity of ALP in the serum at all doses of the extract. This may be an indicative of intra-hepatic cholestasis and pathological condition⁴¹. Also, alkaline phosphatase is a marker of obstructive jaundice or intra-hepatic cholestasis⁴². The bile duct obstruction induces synthesis of this enzyme by biliary tract epithelial cells, leading to very high level of the enzyme in blood circulation⁴¹. The response of the liver to any form of biliary tract obstruction is to induce synthesis of ALP and drugs have been known to cause intra-hepatic obstruction of bile flow^{37,33}. Furthermore, mild elevation of ALP is seen in parenchymal diseases of the liver caused by infectious or toxic hepatitis, due to the effect of drugs or xenobiotics⁴³.

Total protein is composed of albumin and globulin and reflects the balance of protein biosynthesis and catabolism⁴⁴. The increase in serum total protein concentration following the administration of the aqueous extract of *Khaya senegalensis* stem bark could be as a result of tissue damage⁴⁵. Adedapo *et al*⁴⁶ find out that the effect of *K. senegalensis* alteration on the total protein is usually due to decrease in quantity of albumin which may be accompanied by an increase in level of globulin.

The concentration of serum albumin showed a dose dependent decrease when compared with the control. Albumin is preferred to assay for the synthetic function of liver⁴⁷. The decrease in albumin may lead to loss of the integrity of plasma membrane and endoplasmic reticulum⁴⁰. Also, the decrease in the level of albumin is a sign of progressive liver failure and may result in the alteration of total protein and as well as increase level of globulin⁴⁶. The decrease level of albumin will lead to lowering of albumin/globulin ratio (A/G ratio).

The level of globulin was found to show a significant increase at all doses of the extract when compared with the control. The increase in level of globulin may suggest that the extract has the potential to boost the immune system by promotion production and immunoglobulins⁴⁸. The increase in globulin will lead to lowering of albumin/globulin ration (A/G ratio). This increase in globulin will lead to decrease in albumin which will lead to reduced synthetic capacity of the liver.

CONCLUSION

The administration of aqueous extract of *Khaya senegalensis* stem bark has adverse effects on some parameters of liver function of the animals at the doses investigated in this study. Further studies are required for long-term and multiple organs should be involved to examine the effect of dose and duration of consumption of aqueous extract of *Khaya senegalensis* bark stem on the biological system.

REFERENCES

1. Tiwari, U. Rastogi, U. Singh, P. Sharaf, D.K. and Vyas, S.P., Immunomodulatory effects of aqueous extract of *Tridax procumbens* in experimental animals. *J. Ethnopharmacol.* **92**: 113 – 119 (2004)
2. Shafaei, A., T.F., Parsi B.M.K., Ahmad, M.J.A. Siddique, I.H. Italla A.H., Zharia I. and M.Z. Asmawi., Evaluation of Toxicological and standardization parameters and phytochemical investigation of ficus deltoidea leaves. *Am. J. Biochem. Mol. Biol.*, **1**: 237 – 243 (2011)
3. Nevin, K.G and P.L. Vijayammal., Pharmacological and Immunomodulatory effects of Aervalanta in Daltons lymphoma Ascites – Bearing mice. *Pharmacological Biol*, **43**: 640 – 646 (2005)
4. Akpanabiatu, M.I. Umoh, I.B. Udosen, E.O. Udosen, A.E. Udoh A.E. and Edee E.E., Rat serum electrolytes, lipid profile and cardiovascular activity on Nauclea Catifolia leaf extract administration. *Ind. J. Clin. Biochem*, **20**: 29 – 34 (2005)

5. Aboderin, F.I. and V.O. Oyetayo., Hematological Studies of rats feed different doses of Probiotic, *Lactobacillus plantarum*, isolated from fermenting corn slurry. *Park. J. Nutr.* **5**: 102 – 105 (2006)
6. Cheesbrough, M., *Medical Laboratory manual for Tropical Countries*. 2nd edn, Tropical Health Technology and Butter worth scientific Ltd., Cambridge & Edinburgh. **1**: 494 – 526 (1991)
7. Belem, B. Olsen, C.S. Theilade, I. Bellefontaine, R. Guinko, S. Lyk, A.M. Diallo, A. and Boussim, J.I., Identification des arbres hors forêt préférés des populations du Sanmatenga (Burkina Faso). *Bois et forêts des tropiques*, **298** (4): 53-64 (2008)
8. Lompo, M., Etude phamaco-toxicologique chez la souris et le rat de *Khaya senegalensis* (Desr.) A. Juss (Meliaceae) utilisé en tradithérapeutique au Burkina Faso. Mémoire de D.E.A. Physiol. Anim. Appl. Université de Ouagadougou. (1993)
9. Yuan, T. Zhang, C-R. Yang, S.P. and Yue, J-M., Limonoids and Triterpenoids from *Khaya senegalensis*. *J. Nat. Prod.*, **73**: 669–674. (2010)
10. Bickii, J. Njifutie, N. Foyere, J.A. Basco, L.K. and Ringwald, P. *In vitro* antimalarial activity of limonoids from *Khaya grandifoliola* C.D.C. (Meliaceae). *J. Ethnopharmacol.*, **69**: 27–33 (2000)
11. Gill, L.S., *Ethnomedicinal uses of plants in Nigerian Uniben Press*, Benin City Pp. 15 – 65 (1992)
12. Nwosu, C.U. Hassan, S.W. Abubakar, M.G. and Ebbo, A.A., Anti-diarrhoeal and toxicological studies of leaf extracts of *Khaya senegalensis*. *J. Pharm. Toxicol.*, **7**(1): 1-10. (2012)
13. Konaté, K. Kiendrébéogo, M. Ouattara, M.B. Souza, A. Lamien-Meda, A. Nongasida, Y. Barro, N. Millogo-Rasolodimby, J. and Nacoulma, O.G., Antibacterial potential of aqueous acetone extracts from five medicinal plants used traditionally to treat Infectious Diseases in Burkina Faso. *Curr. Res. J. Biol. Sci.*, **3**(5): 435 – 442. (2011)
14. Zhang, H. Wang, X. Chen, F. Androulakis, X.M. and Wargovich, M.J., Anticancer activity of Limonoids from *Khaya senegalensis*. *Phytoter. Res.*, **21**: 731-734. (2007)
15. Androulakis, X.M. Muga, S.J. Chen, F. Koïta, Y. Touré, B. and Wargovich, M.J., Chemopreventive Effects of *Khaya senegalensis* Bark Extract on Human Colorectal Cancer. *Anticancer Res.*, **26**: 2397-2406 (2006)
16. Chiezey, N.P. Gefu, J.O. Jagun, A.G. Abdu, P.A. Alawa, C.B.I. Magaji, S.O. Adeyinka, I.A. Eduvie, L.O., Evaluation of some Nigerian plants for anthelmintic activity in young cattle. Proceedings of the International Workshop on Ethnoveterinary Practices, 14-18 August, Kaduna, Nigeria (2000)
17. Okpara, J.O. Anagor, P.O. Okpalia, E.J. Abdullahi, A. and Ahmed, M.S., The anthelmintic efficacy of medicinal herb extracts against gastrointestinal helminths of sheep. Rapport de la 9th Annual Conference of Animal Science Association of Nigeria (ASAN), 13–16 Sept. Ebonyi State University, Nigeria. (2004)
18. Ibrahim, M.A. Njoku, G.C. and Sallau, A.B., *In vivo* activity of stem barks aqueous extract of *Khaya senegalensis* against *Trypanosoma brucei*. *Afr. J. Biotechnol.*, **7**: 661- 663. (2008)
19. Umar, I.A. Ibrahim, M.A. Fari, N.A. Isah, S. and Balogun, D.A. *In-vitro* and *-vivo* anti-*Trypanosoma evansi* activities of extracts from different parts of *Khaya senegalensis*. *J. Cell Ani. Bio.* **4**(6): 91-95. (2010)
20. Shaba, P. Pandey, N.N. Sharma, O.P. Rao, J.R. Mishra, A.K. and Singh, R.K., Antitrypanosomal activity of methanolic extract of *Khaya Senegalensis* tree bark against *Trypanosoma evansi*. *Int. J. of Food, Agri. And Vet. Sc.*, **1**(1) : 21-26. (2011)
21. Kolawole, O.T. Kolawole, S.O. Ayankunle, A.A. and Olaniran, O.I., Anti-hyperglycemic Effect of *Khaya senegalensis* Stem Bark Aqueous Extract in Wistar Rats. *European J. of Medicinal Plants*, **2**(1): 66-73. (2012)
22. Funke, I. and Melzig, M.F., Traditionally used plants in diabetes therapy - phytotherapeutics as inhibitors of aamylase activity. *Braz. J. Pharmacogn.*, **16**(1): 1-5 (2006)
23. Maydell, H.J., Trees and shrubs of sahel – their characteristics and uses. Gesdesehraye, Fur, Germany, Pp. 105 – 110. (1986)

24. Musa, T.Y. Adebayo, O.J., Egwin, E.C. and Owoyede, V.B., Increased liver alkaline phosphatase and aminotransferases activities following administration of ethanolic extract of *K. senegalensis* stem bark on rats. *Biochem* **17 (1)**: 27 – 32 (2005)
25. Yakubu, M.T. Akanji, M.A. Oladiji, A.T., Aphrodisiac potentials of aqueous extract of *Fadogia agrestis* (Schweinf. Ex Heirn) stem in male albino rats. *Asian J Androl.* **7**:399–404 (2005)
26. Henry, R.J. Cannon, D.C. and Winkelman, J.W., *Clinical Chemistry, Principles and Techniques.* Harper and Row, 2nd ed. (1974)
27. Schmidt, E., Schmidt, F.W., Determination of serum GOT and GPT activities. *Enzyme Biology Chemistry.* **3**:1-5. (1963)
28. Wright, P.J., Leathwood, A.O., Plummer, D.T., Enzymes in rat urine: Alkaline Phosphatase. *Enzymologia*; **42**: 317 – 327. (1972)
29. Grant, G. H. and Kacchman, J. F., In: “Fundamental of Clinical Chemistry” (Tietz, N.W), 3rd edition. W.B. Saunders Company, Philadelphia. pp.298-320 (1987)
30. El Nahhal, Y., Contamination and safety status of plant and food in Arab countries. *J. Appl. Sci.* **4**: 411 – 417 (2004)
31. Adebayo, A.H. Zeng, G.Z. Fan, J.T. Ji, C.J. He, W.J. Xu, J.J. Zhang, Y.M. Akindahunsi, A.A. Kela, R. Tan, N.H., Biochemical, haematological and histological studies of extract of *Ageratum conyzoides* L. in Sprague Dawley rats. *J. Med. Plants Res.* **4(21)**: 2264-2272 (2010)
32. Pramyothin, P., Samosorn, P., Pongshompoo, S., Chaichantipyuth, C., The protective effects of *Phyllanthus emblica* Linn. Extract on ethanol induced rat hepatic injury. *J Ethnopharmacol.* **107(3)**: 361–364. (2006)
33. Wannang, N.N. Jimam, N.S. Omale, S. Maxwell, L.P.D. Steven, S.G. and Aguiyi, J.C., Effect of *Cucumis metuliferus* (*Cucurbitaceae*) fruits on enzymes and haematological parameters in albino rats. *Afr. J. Biotechnol.* **6(22)**: 2515-2518. (2007)
34. Hayes, P.C. Simpson, K.J. and Garden, O.J., Liver and biliary tract disease. In: Davidson’s principles and practice of medicine. 19th ed. **18**: 832-837 (2002)
35. Benjamin, M.N., *Outline of veterinary Clinical pathology*, University Press, IOWA. USA. pp. 229-232 (1978)
36. Ellis, G., Goldberg, D.M. and Spooner, R.J., Serum Enzyme tests in diseases of the liver and biliary tree. *Am. J. Clin. Pathol.* **70**: 248-258 (1978)
37. Burtis, C.A. Ashwood, E.R., Enzymes. In: Tietz Fundamentals of clinical chemistry, 5th ed. W.B. Saunders Company, New York, pp. 352-369 (2001)
38. Wright, P.J. and Plummer, D.T., The use of urinary enzyme measurement to detect renal damages caused by nephrotic compounds. *Biochem. Pharmacol.* **12**: 65. (1974)
39. Shahjahan, M. Sabitha, K.E. Malbika, J. and Shyamala-Devis, C.S., Effect of *Solanum trilobatum* against carbon tetrachloride induced hepatic damage in albino rats. *Indian. J. Med. Res*; **120**: 194-198 (2004)
40. Akanji, M.A. Olagoke, O.A. and Oloyede, O.B., Effect of chronic consumption of metabisulphite on the integrity of rat liver cellular system. *Toxicology*, **81**: 173 – 179 (1993)
41. Ogbe, R.J. Abu, A.H. Eustace, B.B. and Ochalefu, D.O., Safety evaluation of hydroalcoholic extract of *Cochlospermum planchonii* rhizome in rats. *African Journal of Biotechnology.* **10(66)**, pp. 15006-15010 (2011)
42. Davern, T.J. Scharschmidt, B.F., Biochemical liver tests. In: Sleisenger and Fordtran’s Gastrointestinal and liver disease: pathophysiology, diagnosis, management. Feldman M, Friedman LS, Sleisenger MH (eds), 7th ed. Saunders, Philadelphia, pp. 1227 - 1228 (2002)
43. Vasudevan, D.M. Sreekumari, S., Isoenzymes and clinical enzymology. In: Textbook biochemistry for medical students. 5th ed. Jaypee brothers *Med. Pub.* New Delhi. pp. 52-58. (2007)
44. Salau, A.K. Yakubu, M.T. Oluleye, D.S. Oloyede, O.B. Akanji, M.A., Toxicological evaluation of aqueous leaf extract of *Ficus exasperata* on selected tissues of normal Wistar rats. *Centrepont Journal* (Science Edition). **18 (1)**, 55 – 66 (2012)

45. Aliyu, Donatin R.H. and Jaryum K.H., The Effects of *Boswellia daziellii* on aqueous bark extract on rat liver function. *Asian J. Biochem.*, **2**; 359 – 363 (2007)
46. Adedapo, A.D.A. Osude, Y.O. Adedapo, A.A. Moody, J.O. Adeagbo, A.S. Olajide, O.A. and Makinde, J.M., Blood pressure lowering of *Adenanthera pavinona* seed extract on normotensive rats. *Rec. Nat. Prod.*, **3**: 82 – 89 (2009)
47. Corless, J., and Middleton, A., Normal liver function, a basis for understanding hepatic disease. *Arch. Internled.* **143**: 2291 – 2294 (1993)
48. Puri, A.R. Sakena, R.P. Sarena, K.C., Saxena, Strivastar and J.S., Tandon Immunostimulant agents from *Andrographis paniculata*. *J. Nat. Prod.*, **58**: 998 – 999. (1993)